

### **PCT**

#### **NOTIFICATION OF ELECTION**

(PCT Rule 61.2)

### From the INTERNATIONAL BUREAU

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Applicant

CLARK, Susan, Joy et al

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A. CLASSIFICATION OF SUBJECT MATTER
IPC 6 C12Q1/68 C12N9/10

A61K48/00

G01N33/577

According to International Patent Classification (IPC) or to both national classification and IPC

## B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols) IPC 6 C12Q C12N A61K G01N

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

C. DOCU	MENTS CONSIDERED TO BE RELEVANT	
Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	PROC. NATL.ACAD SCI., vol. 85, no. 14, September 1988 NATL. ACAD SCI.,WASHINGTON,DC,US;, pages 6518-6522, J.A. MOSCOW ET AL. 'Isolation of the human anionic glutathione S-transferase cDNA and the reaction of its gene expression to estrogen-receptor content in primary breast cancer' see the whole document	24
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Further documents are listed in the continuation of box C.	X Patent family members are listed in annex.
* Special categories of cited documents:  *A* document defining the general state of the art which is not considered to be of particular relevance  *E* earlier document but published on or after the international filing date  *U* document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)  *O* document referring to an oral disclosure, use, exhibition or other means  *P* document published prior to the international filing date but later than the priority date claimed	"T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention.  "X" document of particular relevance; the claimed invention carnot be considered howd or cannot be considered to involve an inventive step when the document is taken alone.  "Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination teing obvious to a person skilled in the art.  "&" document member of the same patent family
Date of the actual completion of the international search  7 November 1995	Date of mailing of the international search report  3 0. 11, 95
Name and mailing address of the ISA  European Patent Office, P.B. SEE Patentiaan 2 NL - 2280 HV Rijswijk Tel. (+31-70) 340-2040, Tk. 31-651 epo nl.	Authorized officer

# INTERNATIONAL SEARCH REPORT

are Roux	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
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<b>A</b>	J UROL 150 (1). 1993. 209-214. CODEN: JOURAA ISSN: 0022-5347, July 1993 RIPPLE M ET AL 'CHARACTERISTICS OF THE GLUTATHIONE GLUTATHIONE - S - TRANSFERASE DETOXIFICATION SYSTEM IN MELPHALAN RESISTANT HUMAN PROSTATE CANCER CELLS.' see the whole document	1-30
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C (Conunu	IUON) DOCUMENTS CONSIDERED TO BE RELEVANT	
Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A	85TH ANNUAL MEETING OF THE AMERICAN ASSOCIATION FOR CANCER RESEARCH, SAN FRANCISCO, CALIFORNIA, USA, APRIL 10-13, 1994. PROCEEDINGS OF THE AMERICAN ASSOCIATION FOR CANCER RESEARCH ANNUAL MEETING 35 (0). 1994. 293. ISSN: 0197-016X, ABSTRACT NO. 1746, BUETOW K H ET AL 'GSTM1 and risk for breast and prostate cancer.' see abstract	1-30
<b>A</b>	EIGHTY-EIGHTH ANNUAL MEETING OF THE AUA (AMERICAN UROLOGICAL ASSOCIATION), SAN ANTONIO, TEXAS, USA, MAY 15-20, 1993. J UROL 149 (4 SUPPL.). 1993. 470A. CODEN: JOURAA ISSN: 0022-5347, ABSTRACT NO.1028, BROOKS J D ET AL 'ALTERED REGULATION OF GLUTATHIONE - S - TRANSFERASE PI IN HUMAN PROSTATIC CANCER CELLS.' see abstract	1-30
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C (Continu	Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT				
'ategory "	Citation of document, with indication, where appropriate, of the relevant passages		Relevant to claim No.		
P,X	NINETIETH ANNUAL MEETING OF THE AMERICAN UROLOGICAL ASSOCIATION, LAS VEGAS, NEVADA, USA, APRIL 23-28, 1995. JOURNAL OF UROLOGY 153 (4 SUPPL.). 1995. 446A. ISSN: 0022-5347, ABSTRACT NO. 869, LEE W-H ET AL 'Cytidine methylation of the pi-class glutathione S - transferase gene promoter in human prostate cancer.' see abstract		1-30		
P,X	EIGHTY-SIXTH ANNUAL MEETING OF THE AMERICAN ASSOCIATION FOR CANCER RESEARCH, TORONTO, ONTARIO, CANADA, MARCH 18-22, 1995. PROCEEDINGS OF THE AMERICAN ASSOCIATION FOR CANCER RESEARCH ANNUAL MEETING 36 (0). 1995. 538. ISSN: 0197-016X, ABSTRACT NO. 3206, LEE W-H ET AL 'Methylation of 5' regulatory sequences near the GSTP1 gene in human prostatic carcinomas.' see abstract		1-30		



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Box 1 Observations where certain claims were found unsearchable (Continuation of item 1 of first sheet)

This international search report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:
Claims Nos.: because they relate to subject matter not required to be searched by this Authority, namely:  Remark: Although claims 19-23,27 and 24-26,28-30 (partially),(as far as in vivo methods are concerned) are directed to a method of treatment of the human/animal body, the search has been carried out and based on the alleged effects of the compound/composition.
Claims Nos.:  because they relate to parts of the international application that do not comply with the prescribed requirements to such an extent that no meaningful international search can be carried out, specifically:
3. Claims Nos.: because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).
Box II Observations where unity of invention is lacking (Continuation of item 2 of first sheet)
This International Searching Authority found multiple inventions in this international application, as follows.
As all required additional search fees were timely paid by the applicant, this international search report covers all searchable claims.
2. As all searchable claims could be searches without effort justifying an additional fee, this Authority did not invite payment of any additional fee.
As only some of the required additional search fees were timely paid by the applicant, this international search report covers only those claims for which fees were paid, specifically claims Nos
No required additional search fees were timely paid by the applicant. Consequently, this international search report is restricted to the invention first mentioned in the claims; it is covered by claims Nos

The additional search fees were accompanied by the applicant s protest



INTERNATIONAL SEARCH REPORT

Patent document cited in search report	Publication date	Patent family member(s)		Publication date	
WO-A-9012088	18-10-90	AU-B-	5435990	05-11-90	

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said site or sites at which abnormal cytosine methylation occurs is/are methylated, and

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(iii) determining the presence of amplified DNA.

Since the amplification is designed to only amplify the target region if the said site or sites at which abnormal cytosine methylation (i.e. as compared to the corresponding site or sites of DNA from subjects without the disease or condition being assayed) occurs is/are methylated, the presence of amplified DNA will be indicative of the disease or condition in the subject from which the isolated DNA has been obtained. The assay thereby provides a means for diagnosing or prognosing the disease or condition in a subject.

The step of isolating DNA may be conducted in accordance with standard protocols. The DNA may be isolated from any suitable body sample, such as cells from tissue (fresh or fixed samples), blood (including serum and plasma), semen, urine, lymph or bone marrow. For some types of body samples, particularly fluid samples such as blood, semen, urine and lymph, it may be preferred to firstly subject the sample to a process to enrich the concentration of a certain cell type (e.g. prostate cells). One suitable process for enrichment involves the separation of required cells through the use of cell-specific antibodies coupled to magnetic beads and a magnetic cell separation device.

Prior to the amplifying step, the isolated DNA is preferably treated such that unmethylated cytosines are converted to uracil or another nucleotide capable of forming a base pair with adenine while methylated cytosines are unchanged or are converted to a nucleotide capable of forming a base pair with guanine. This treatment permits the design of primers which enable the selective amplification of the target region if the said site or sites at which abnormal cytosine methylation occurs is/are methylated.

Preferably, following treatment and amplification of the isolated DNA, a test is performed to verify that unmethylated cytosines have been efficiently converted to uracil or another nucleotide capable of forming a

base pair with adenine, and that methylated cystosines have remained unchanged or efficiently converted to another nucleotide capable of forming a base pair with guanine.

Preferably, the treatment of the isolated DNA involves reacting the isolated DNA with bisulphite in accordance with standard protocols. As will be clear from the above discussion of bisulphite treatment, unmethylated cytosines will be converted to uracil whereas methylated cytosines will be unchanged. Verification that unmethylated cytosines have been converted to uracil and that methylated cystosines have remained unchanged may be achieved by:

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- (i) restricting an aliquot of the treated and amplified DNA with a suitable restriction enzyme(s) which recognise a restriction site(s) generated by or resistant to the bisulphite treatment, and
- (ii) assessing the restriction fragment pattern by electrophoresis.

  Alternatively, verification may be achieved by differential hybridisation using specific oligonucleotides targeted to regions of the treated DNA where unmethylated cytosines would have been converted to uracil and methylated cytosines would have remained unchanged.

The amplifying step may involve polymerase chain reaction (PCR) amplification, ligase chain reaction amplification (20) and others (21).

Preferably, the amplifying step is conducted in accordance with standard protocols for PCR amplification, in which case, the reactants will typically be suitable primers, dNTPs and a thermostable DNA polymerase, and the conditions will be cycles of varying temperatures and durations to effect alternating denaturation of strand duplexes, annealing of primers (e.g. under high stringency conditions) and subsequent DNA synthesis.

To achieve selective PCR amplification with bisulphite-treated DNA, primers and conditions may be used to discriminate between a target region including a site or sites of abnormal cytosine methylation and a target region where there is no site or sites of abnormal cytosine methylation. Thus, for

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amplification only of a target region where the said site or sites at which abnormal cytosine methylation occurs is/are methylated, the primers used to anneal to the bisulphite-treated DNA (i.e. reverse primers) will include a guanine nucleotide(s) at a site(s) at which it will form a base pair with a methylated cytosine(s). Such primers will form a mismatch if the target region in the isolated DNA has unmethylated cytosine nucleotide(s) (which would have been converted to uracil by the bisulphite treatment) at the site or sites at which abnormal cytosine methylation occurs. The primers used for annealing to the opposite strand (i.e. the forward primers) will include a cytosine nucleotide(s) at any site(s) corresponding to site(s) of methylated cytosine in the bisulphite-treated DNA.

Preferably, the primers used for the PCR amplification are of 12 to 30 nucleotides in length and are designed to anneal to a sequence within the target region that includes two to four cytosine nucleotides that are abnormally methylated in the DNA of a subject with the disease or condition being assayed. In addition, the primers preferably include a terminal nucleotide that will form a base pair with a cytosine nucleotide (reverse primer), or the guanine nucleotide opposite (forward primer), that is abnormally methylated in the DNA of a subject with the disease or condition being assayed.

The step of amplifying is used to amplify a target region within the GST-Pi gene and/or its regulatory flanking sequences. The regulatory flanking sequences may be regarded as the flanking sequences 5' and 3' of the GST-Pi gene which include the elements that regulate, either alone or in combination with another like element, expression of the GST-Pi gene. Preferably, the regulatory flanking sequences consist of the 400 nucleotide sequence immediately 5' of the transcription start site and the 100 nucleotide sequence immediately 3' of the transcription stop site.

More preferably, the step of amplifying is used to amplify a target region within the region of the GST-Pi gene and its regulatory flanking

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sequences defined by (and inclusive of) CpG sites -43 to +55 (wherein the numbering of the CpG sites is relative to the transcription start site). The numbering and position of CpG sites is shown in Figure 1.

The step of determining the presence amplified DNA may be conducted in accordance with standard protocols. One convenient method involves visualisation of a band(s) corresponding to amplified DNA, following gel electrophoresis.

Preferably, the disease or condition to be assayed is selected from cancers, especially hormone dependent cancers such as prostate cancer, breast cancer and cervical cancer, and liver cancer.

For the diagnosis or prognosis of prostate cancer, the step of amplifying preferably amplifies a target region within the region of the GST-Pi gene and its regulatory flanking sequences defined by (and inclusive of) CpG sites -43 to +53, more preferably, -43 to +10. However, within these target regions it is believed that there are CpG sites which show variability in methylation status in prostate cancer or are methylated in other tissues. Thus, for the target region defined by (and inclusive of) CpG sites -43 to +10, it is preferred that the primers used for amplification be designed so as to minimise (i.e. by use of redundant primers or by avoidance of the sites) the influence of CpG sites -36, -32, -23, -20, -14 and a polymorphic region covering site -33. Further, for DNA isolated from cells other than from prostate tissue (e.g. blood), it is preferred that the primers used be designated to amplify a target region that does not include the region of the GST-Pi gene and its regulatory flanking sequences defined by (and inclusive of) CpG sites -7 to +7, or, more preferably, -13 to +8, since this may lead to false positives. Further preferred target regions, therefore, are within the region of the GST-Pi gene and its regulatory flanking sequences defined by (and inclusive of) CpG sites -43 to -14, -43 to -8, +9 to +53 and +1 to +53.

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Suitable primer pairs for the diagnosis or prognosis of prostate cancer, include those consisting of a forward and reverse primer selected from each of the following groups:

	Forward Primers (i.e. anneal to the 5' end of the targe	et region)
5	CGCGAGGTTTTCGTTGGAGTTTCGTCGTC	(SEQ ID NO: 1)
	CGTTATTAGTGAGTACGCGCGGTTC	(SEQ ID NO: 2)
	YGGTTTTAGGGAATTTTTTTCGC	(SEQ ID NO: 3)
	YGGYGYGTTAGTTYGTTGYGTATATTTC	(SEQ ID NO: 4)
	GGGAATTTTTTTCGCGATGTTTYGGCGC	(SEQ ID NO: 5)
10	TTTTAGGGGGTTYGGAGCGTTTC	(SEQ ID NO: 6)
	GGTAGGTTGYGTTTATCGC	(SEQ ID NO: 7)
	Reverse Primers (i.e. anneal to the extension of the fo	orward primer)
	TCCCATCCCTCCCGAAACGCTCCG	(SEQ ID NO: 8)
	GAAACGCTCCGAACCCCCTAAAAACCGCTAACG	(SEQ ID NO: 9)
15	CRCCCTAAAATCCCCRAAATCRCCGCG	(SEQ ID NO: 10)
	ACCCCRACRACCRCTACACCCCRAACGTCG	(SEQ ID NO: 11)
	CTCTTCTAAAAAATCCCRCRAACTCCCGCCG	(SEQ ID NO: 12)
	AAAACRCCCTAAAATCCCCGAAATCGCCG	(SEQ ID NO: 13)
	AACTCCCRCCGACCCCAACCCCGACGACCG	(SEQ ID NO: 14)
20	AAAAATTCRAATCTCTCCGAATAAACG	(SEQ ID NO: 15)
	AAAAACCRAAATAAAAACCACACGACG	(SEQ ID NO: 16)
	wherein Y is C, T or, preferably, a mixture thereof, and R is	s A, G or,
	preferably, a mixture thereof.	

For the diagnosis or prognosis of liver cancer, the step of amplifying preferably amplifies a target region within the region of the GST-Pi gene and its regulatory flanking sequences defined by (and inclusive of) CpG sites -43 to -14 and/or +9 to +53. However, within these target regions it is believed that there are CpG sites which show variability in methylation status in liver cancer or are methylated in other tissues. Thus, for the target region defined by (and inclusive of) CpG sites -43 to -14, it is preferred that the primers used

for amplification be designed so as to minimise (i.e. by use of redundant primers or by avoidance of the sites) the influence of CpG sites -36, -32, -23, -20, -14 and a polymorphic region covering site -33.

It will be appreciated by persons skilled in the art, that a site or sites of abnormal cytosine methylation within the above identified target regions of the GST-Pi gene and/or its regulatory flanking sequences, could be detected for the purposes of diagnosing or prognosing a disease or condition (particularly, prostate cancer and/or liver cancer) by methods which do not involve selective amplification. For instance, oligonucleotide/polynucleotide probes could be designed for use in hybridisation studies (e.g. Southern blotting) with bisulphite-treated DNA which, under appropriate conditions of stringency, selectively hybridise only to DNA which includes a site or sites of abnormal methylation of cytosine(s). Alternatively, an appropriately selected informative restriction enzyme(s) could be used to produce restriction fragment patterns that distinguish between DNA which does and does not include a site or sites of abnormal methylation of cytosine(s).

Thus, in a second aspect, the present invention provides a diagnostic or prognostic assay for a disease or condition in a subject said disease or condition characterised by abnormal methylation of cytosine at a site or sites within the glutathione-S-transferase (GST) Pi gene and/or its regulatory flanking sequences, wherein said assay comprises the steps of;

(i) isolating DNA from said subject, and

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(ii) determining the presence of abnormal methylation of cytosine at a site or sites within the region of the GST-Pi gene and/or its regulatory flanking sequences defined by (and inclusive of) CpG sites -43 to +55.

The step of isolating DNA may be conducted as described above in relation to the assay of the first aspect.

Preferably, the region of the GST-Pi gene and its regulatory flanking sequences within which the presence of methylated cytosine(s) at a site or sites is determined is selected from the regions defined by (and inclusive of)

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CpG sites -43 to +53, -43 to +10, -43 to -14, +9 to +53 and +1 to +53. However, within these regions, it is preferred that certain sites (namely, CpG sites, -36, -33, -32, -23, -20, -17 and -14) be avoided as the site or sites at which, for the purpose of the assay, the presence of abnormal methylation of cytosine is determined.

Where the determination step is to involve selective hybridisation of oligonucleotide/polynucleotide/peptide-nucleic acid (PNA) probes, prior to the determination step, the isolated DNA is preferably treated (e.g. with bisulphite) such that unmethylated cytosines are converted to uracil or another nucleotide capable of forming a base pair with adenine while methylated cytosines are unchanged or are converted to a nucleotide capable of forming a base pair with guanine. This treatment permits the design of probes which allow for selective hybridisation to a target region including a site or sites of abnormal methylation of cytosine.

In a third aspect, the present invention provides a primer or probe (sequence shown in the 5' to 3' direction) comprising a nucleotide sequence selected from the group consisting of:

	CGCGAGGTTTCGTTGGAGTTTCGTCGTC	(SEQ ID NO: 1)
	CGTTATTAGTGAGTACGCGCGGTTC	(SEQ ID NO: 2)
20	YGGTTTTAGGGAATTTTTTTCGC	(SEQ ID NO: 3)
	YGGYGYGTTAGTTYGTTGYGTATATTTC	(SEQ ID NO: 4)
	GGGAATTTTTTTCGCGATGTTTYGGCGC	(SEQ ID NO: 5)
	TTTTTAGGGGGTTYGGAGCGTTTC	(SEQ ID NO: 6)
	GGTAGGTTGYGTTTATCGC	(SEQ ID NO: 7)
25	AAAAATTCRAATCTCTCCGAATAAACG	(SEQ ID NO: 8)
	AAAAACCRAAATAAAAACCACACGACG	(SEQ ID NO: 9)
	TCCCATCCCTCCCGAAACGCTCCG	(SEQ ID NO: 10)
	GAAACGCTCCGAACCCCCTAAAAACCGCTAACG	(SEQ ID NO: 11)
	CRCCCTAAAATCCCCRAAATCRCCGCG	(SEQ ID NO: 12)
<b>3</b> 0	ACCCCRACRACCRCTACACCCCRAACGTCG	(SEQ ID NO: 13)



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### Claims:

- 1. A diagnostic or prognostic assay for a disease or condition in a subject, said disease or condition characterised by abnormal methylation of cytosine at a site or sites within the glutathione-S-transferase (GST) Pi gene and/or its regulatory flanking sequences, wherein said assay comprises the steps of;
- (i) isolating DNA from said subject,
- (ii) exposing said isolated DNA to reactants and conditions for the
  amplification of a target region of the GST-Pi gene and/or its regulatory
  flanking sequences which includes a site or sites at which abnormal cytosine
  methylation characteristic of the disease or condition occurs, the
  amplification being selective in that it only amplifies the target region if the
  said site or sites at which abnormal cytosine methylation occurs is/are
  methylated, and
  - (iii) determining the presence of amplified DNA.
  - 2. An assay according to claim 1, wherein the amplifying step is used to amplify a target region within the GST-Pi gene and/or its regulatory flanking sequences, wherein the regulatory flanking sequences consist of the 400 nucleotide sequence immediately 5' of the transcription start site of the GST-Pi gene and the 100 nucleotide sequence immediately 3' of the transcription stop site of the GST-Pi gene.
- 25 3. An assay according to claim 1 or 2, wherein the amplifying step is used to amplify a target region within the region of the GST-Pi gene and its regulatory flanking sequences defined by (and inclusive of) CpG sites -43 to +55.
- 4. An assay according to any one of the preceding claims, wherein prior to the amplifying step, the isolated DNA is treated such that unmethylated

cytosines are converted to uracil or another nucleotide capable of forming a base pair with adenine while methylated cytosines are unchanged or are converted to a nucleotide capable of forming a base pair with guanine.

- 5. As assay according to any one of the preceding claims, wherein the amplifying step involves polymerase chain reaction (PCR) amplification.
  - 6. An assay according to claim 5, wherein said PCR amplification utilises a reverse primer including guanine at at least one site whereby, upon the reverse primer annealing to the treated DNA, said guanine will either form a base pair with a methylated cytosine (or another nucleotide to which the methylated cytosine has been converted through said treatment) if present, or will form a mismatch with uracil (or another nucleotide to which unmethylated cytosine has been converted through said treatment).

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7. An assay according to claim 6, wherein said PCR amplification utilises a forward primer including cytosine at at least one site(s) corresponding to cytosine nucleotides that are abnormally methylated in the DNA of a subject with the disease or condition being assayed.

- 8. An assay according to claim 7, wherein the primers are of 12 to 30 nucleotides in length.
- 9. An assay according to claim 8, wherein the primers are selected so as to anneal to a sequence within the target region that includes two to four cytosine nucleotides that are abnormally methylated in the isolated DNA of a subject with the disease or condition being assayed.
- An assay according to claim 4, wherein the treatment of the isolated
   DNA involves reacting the isolated DNA with bisulphite.

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- 11. As assay according to claim 10, wherein the amplifying step involves polymerase chain reaction (PCR) amplification.
- An assay according to claim 11, wherein said PCR amplification utilises a reverse primer including guanine at at least one site whereby, upon the reverse primer annealing to the treated DNA, said guanine will either form a base pair with a methylated cytosine if present, or will form a mismatch with uracil.

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13. An assay according to claim 12, wherein said PCR amplification utilises a forward primer including cytosine at at least one site(s) corresponding to cytosine nucleotides that are abnormally methylated in the isolated DNA of a subject with the disease or condition being assayed.

- 14. An assay according to claim 13, wherein the primers are of 12 to 30 nucleotides in length.
- 15. An assay according to claim 14, wherein the primers are selected so as to anneal to a sequence within the target region that includes two to four cytosine nucleotides that are abnormally methylated in the DNA of a subject with the disease or condition being assayed.
- An assay according to any one of the preceding claims, wherein said
   DNA is isolated from cells from tissue, blood (including serum and plasma),
   semen, urine, lymph or bone marrow.
  - 17. An assay according to any one of the preceding claims, wherein the disease or condition to be assayed is selected from cancers.

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- 18. An assay according to claim 17, wherein the disease or condition to be assayed is selected from prostate cancer, breast cancer, cervical cancer and liver cancer.
- 5 19. An assay according to claim 18, wherein the disease or condition to be assayed is prostate cancer.
  - 20. An assay according to claim 19, wherein the amplifying step is used to amplify a target region within the region of the GST-Pi gene and its regulatory flanking sequences defined by (and inclusive of) CpG sites -43 to +53.
    - 21. An assay according to claim 19, wherein the amplifying step is used to amplify a target region within the region of the GST-Pi gene and its regulatory flanking sequences defined by (and inclusive of) CpG sites -43 to +10.
  - 22. An assay according to claim 19, wherein the amplifying step is used to amplify a target region within the region of the GST-Pi gene and its regulatory flanking sequences defined by (and inclusive of) CpG sites -43 to -14.
    - 23. An assay according to claim 19, wherein the amplifying step is used to amplify a target region within the region of the GST-Pi gene and its regulatory flanking sequences defined by (and inclusive of) CpG sites -43 to -8.
    - 24. An assay according to claim 19, wherein the amplifying step is used to amplify a target region within the region of the GST-Pi gene and its

regulatory flanking sequences defined by (and inclusive of) CpG sites +9 to +53.

- 25. An assay according to claim 19, wherein the amplifying step is used to amplify a target region within the region of the GST-Pi gene and its regulatory flanking sequences defined by (and inclusive of) CpG sites +1 to +53.
- 26. An assay according to claim 19, wherein the amplifying step involves
  PCR amplification using primer pairs consisting of a foward and reverse
  primer selected from each of the following groups:

	Forward Primers	
	CGCGAGGTTTTCGTTGGAGTTTCGTCGTC	(SEQ ID NO: 1)
	CGTTATTAGTGAGTACGCGCGGTTC	(SEQ ID NO: 2)
15	YGGTTTTAGGGAATTTTTTTCGC	(SEQ ID NO: 3)
	YGGYGYGTTAGTTYGTTGYGTATATTTC	(SEQ ID NO: 4)
	GGGAATTTTTTTCGCGATGTTTYGGCGC	(SEQ ID NO: 5)
	TTTTTAGGGGGTTYGGAGCGTTTC	(SEQ ID NO: 6)
	GGTAGGTTGYGTTTATCGC	(SEQ ID NO: 7)
20	Reverse Primers	
	TCCCATCCCTCCCGAAACGCTCCG	(SEQ ID NO: 8)
	GAAACGCTCCGAACCCCCTAAAAACCGCTAACG	(SEQ ID NO: 9)
	CRCCCTAAAATCCCCRAAATCRCCGCG	(SEQ ID NO: 10)
	ACCCCRACRACCTCTACACCCCRAACGTCG	(SEQ ID NO: 11)
25	CTCTTCTAAAAAATCCCRCRAACTCCCGCCG	(SEQ ID NO: 12)
	AAAACRCCCTAAAATCCCCGAAATCGCCG	(SEQ ID NO: 13)
	AACTCCCRCCGACCCCAACCCCGACGACCG	(SEQ ID NO: 14)
	AAAAATTCRAATCTCTCCGAATAAACG	(SEQ ID NO: 15)
	AAAAACCRAAATAAAAACCACACGACG	(SEQ ID NO: 16),
30	wherein Y is C, T or a mixture thereof, and R is A, G or a n	nixture thereof.

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27. An assay according to claim 19, wherein the amplifying step involves PCR amplification using primer pairs consisting of a foward and reverse primer selected from each of the following groups:

Forward Primers	
CGCGAGGTTTCGTTGGAGTTTCGTCGTC	(SEQ ID NO: 1)
CGTTATTAGTGAGTACGCGCGGTTC	(SEQ ID NO: 2)
Reverse Primers	
TCCCATCCCTCCCGAAACGCTCCG	(SEQ ID NO: 8)
GAAACGCTCCGAACCCCCTAAAAACCGCTAACG	(SEO ID NO: 9)

28. An assay according to claim 19, wherein the amplifying step involves PCR amplification using primer pairs consisting of a foward and reverse primer selected from each of the following groups:

	Forward Primers	
15	YGGTTTTAGGGAATTTTTTTCGC	(SEQ ID NO: 3)
	YGGYGYGTTAGTTYGTTGYGTATATTTC	(SEQ ID NO: 4)
	GGGAATTTTTTTCGCGATGTTTYGGCGC	(SEQ ID NO: 5)
	Reverse Primers	
	CRCCCTAAAATCCCCRAAATCRCCGCG	(SEQ ID NO: 10)
20	ACCCCRACRACCRCTACACCCCRAACGTCG	(SEQ ID NO: 11)
	CTCTTCTAAAAAATCCCRCRAACTCCCGCCG	(SEQ ID NO: 12)
	AAAACRCCCTAAAATCCCCGAAATCGCCG	(SEQ ID NO: 13)
	AACTCCCRCCGACCCCAACCCCGACGACCG	(SEQ ID NO: 14),
	wherein Y is C, T or a mixture thereof and R is A, G or a	mixture thereof.

29. An assay according to claim 19, wherein the amplifying step involves PCR amplification using primer pairs consisting of a forward and reverse

primer selected from each of the following groups:

### Forward Primers

TTTTTAGGGGGTTYGGAGCGTTTC (SEQ ID NO: 6)

GGTAGGTTGYGTTTATCGC (SEQ ID NO: 7)

Reverse Primers

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15

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AAAAATTCRAATCTCTCCGAATAAACG (SEQ ID NO: 15)

AAAAACCRAAATAAAAACCACACGACG (SEQ ID NO: 16),

wherein Y is C, T or a mixture thereof, and R is A, G or a mixture thereof.

- 30. An assay according to claim 18, wherein the disease or condition to be assayed is liver cancer.
  - 31. An assay according to claim 30, wherein the amplifying step is used to amplify a target region within the region of the GST-Pi gene and its regulatory flanking sequences defined by (and inclusive of) CpG sites -43 to -14.
  - 32. An assay according to claim 30, wherein the amplifying step is used to amplify a target region within the region of the GST-Pi gene and its regulatory flanking sequences defined by (and inclusive of) CpG sites +9 to +53.
  - 33. A diagnostic or prognostic assay for a disease or condition in a subject said disease or condition characterised by abnormal methylation of cytosine at a site or sites within the glutathione-S-transferase (GST) Pi gene and/or its regulatory flanking sequences, wherein said assay comprises the steps of;
  - (i) isolating DNA from said subject, and
  - (ii) determining the presence of abnormal methylation of cytosine at a site or sites within the region of the GST-Pi gene and/or its regulatory flanking sequences defined by (and inclusive of) CpG sites -43 to +55.

PCT/AU99/00306

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- 34. An assay according to claim 33, wherein the region of the GST-Pi gene and its regulatory flanking sequences within which the presence of methylated cytosine(s) at a site or sites is determined is selected from the regions defined by (and inclusive of) CpG sites -43 to +53, -43 to +10, -43 to -14, +9 to +53 and +1 to +53.
- 35. An assay according to claim 34, wherein the region of the GST-Pi gene and its regulatory flanking sequences within which the presence of methylated cytosine(s) at a site or sites is determined is the region defined by (and inclusive of) CpG sites +9 to +53.
- 36. An assay according to claim 34, wherein the region of the GST-Pi gene and its regulatory flanking sequences within which the presence of methylated cytosine(s) at a site or sites is determined is the region defined by (and inclusive of) CpG sites +1 to +53.
- 37. An assay according to any one of claims 33 to 36, wherein prior to the determination step, the isolated DNA is treated such that unmethylated cytosines are converted to uracil or another nucleotide capable of forming a base pair with adenine while methylated cytosines are unchanged or are converted to a nucleotide capable of forming a base pair with guanine.
- 38. An assay according to claim 37, wherein the treatment of the isolated DNA involves reacting the isolated DNA with bisulphite.
  - 39. An assay according to any one of claims 33 to 38, wherein the determination step involves selective hybridisation of oligonucleotide/polynucleotide/peptide-nucleic acid (PNA) probes.

- 40. An assay according to any one of claims 33 to 39, wherein said DNA is isolated from cells from tissue, blood (including serum and plasma), semen, urine, lymph or bone marrow.
- 5 41. An assay according to any one of claims 33 to 40, wherein the disease or condition to be assayed is selected from cancers.
  - 42. An assay according to claim 41, wherein the disease or condition to be assayed is selected from prostate cancer, breast cancer, cervical cancer and liver cancer.
    - 43. An assay according to claim 42, wherein the disease or condition to be assayed is prostate cancer.
- 44. An assay according to claim 42, wherein the disease or condition to be assayed is liver cancer.
  - 45. A primer or probe comprising a nucleotide sequence selected from the group consisting of:

20	CGCGAGGTTTCGTTGGAGTTTCGTCGTC	(SEQ ID NO: 1)
	CGTTATTAGTGAGTACGCGCGGTTC	(SEQ ID NO: 2)
	YGGTTTTAGGGAATTTTTTTCGC	(SEQ ID NO: 3)
	YGGYGYGTTAGTTYGTTGYGTATATTTC	(SEQ ID NO: 4)
	GGGAATTTTTTTCGCGATGTTTYGGCGC	(SEQ ID NO: 5)
25	TTTTTAGGGGGTTYGGAGCGTTTC	(SEQ ID NO: 6)
	GGTAGGTTGYGTTTATCGC	(SEQ ID NO: 7)
	AAAAATTCRAATCTCTCCGAATAAACG	(SEQ ID NO: 8)
	AAAAACCRAAATAAAAACCACACGACG	(SEQ ID NO: 9)
	TCCCATCCCTCCCGAAACGCTCCG	(SEQ ID NO: 10)
30	GAAACGCTCCGAACCCCCTAAAAACCGCTAACG	(SEQ ID NO: 11)

CRCCCTAAAATCCCCRAAATCRCCGCG	(SEQ ID NO: 12)
ACCCCRACRACCRCTACACCCCRAACGTCG	(SEQ ID NO: 13)
CTCTTCTAAAAAATCCCRCRAACTCCCGCCG	(SEQ ID NO: 14)
AAAACRCCCTAAAATCCCCGAAATCGCCG	(SEQ ID NO: 15)
AACTCCCRCCGACCCCAACCCCGACGACCG,	(SEQ ID NO: 16),

wherein Y is a mixture of C and T, and R is a mixture of A and G.

- 46. A probe comprising a nucleotide sequence selected from the group consisting of:
- 10 Conversion oligonucleotide:

AAACCTAAAAAATAAACAAACAA (SEQ ID NO: 17)

GGGCCTAGGGAGTAAACAGACAG (SEQ ID NO: 18)

CCTTTCCCTCTTTCCCARRTCCCCA (SEQ ID NO: 19)

TTTGGTATTTTTTTCGGGTTTTAG (SEQ ID NO: 20)

CTTGGCATCCTCCCCGGGCTCCAG (SEQ ID NO: 21)

GGYAGGGAAGGGAGGYAGGGGYTGGG (SEQ ID NO: 22).

15

## **Sequence Listings:**

Applicant: Commonwealth Scientific and Industrial Research

Organisation

Title: Diagnostic assay

Prior Application Number: PP3129

Prior Application Filing Date: 1998-04-23

Number of SEQ ID NOs: 59

Software: PatentIn Ver. 2.1

SEQ ID NO: 1

Length: 29

Type: DNA

Organism: Homo sapiens

Sequence: 1

cgcgaggttt tcgttggagt ttcgtcgtc

SEQ ID NO: 2

Length: 25

Type: DNA

Organism: Homo sapiens

Sequence: 2

cgttattagt gagtacgcgc ggttc

SEQ ID NO: 3 Length: 24

Type: DNA

Organism: Homo sapiens

29

WO 99/55905

PCT/AU99/00306

**4**6

Sequence: 3

yggttttagg gaattttttt tcgc

24

SEQ ID NO: 4 Length: 28

Type: DNA

Organism: Homo sapiens

Sequence: 4

Yggygygtta gttygttgyg tatatttc

28

SEQ ID NO: 5 Length: 29 Type: DNA

Organism: Homo sapiens

Sequence: 5

gggaattttt tttcgcgatg tttyggcgc

29

SEQ ID NO: 6 Length: 24 Type: DNA

Organism: Homo sapiens

Sequence: 6

tttttagggg gttyggagcg tttc

24

SEQ ID NO: 7 Length: 19 Type: DNA

Organism: Homo sapiens

Sequence: 7

ggtaggttgy gtttatcgc

19

SEQ ID NO: 8

47

Length: 27
Type: DNA

Organism: Homo sapiens

Sequence: 8

aaaaattcra atctctccga ataaacg 27

SEQ ID NO: 9 Length: 27

Type: DNA

Organism: Homo sapiens

Sequence: 9

aaaaaccraa ataaaaacca cacgacg 27

SEQ ID NO: 10 Length: 25

Type: DNA

Organism: Homo sapiens

Sequence: 10

tcccatcct ccccgaaacg ctccg 25

SEQ ID NO: 11 Length: 33 Type: DNA

Organism: Homo sapiens

Sequence: 11

gaaacgctcc gaacccccta aaaaccgcta acg 33

SEQ ID NO: 12

Length: 27

Type: DNA

Organism: Homo sapiens

48

Sequence: 12

crecetaaaa teeeeraaat ereegeg 27

SEQ ID NO: 13

Length: 30 Type: DNA

Organism: Homo sapiens

Sequence: 13

accceracra ceretacace ceraaegteg 30

SEQ ID NO: 14

Length: 31
Type: DNA

Organism: Homo sapiens

Sequence: 14

ctcttctaaa aaatcccrcr aactcccgcc g 31

SEQ ID NO: 15

Length: 29
Type: DNA

Organism: Homo sapiens

Sequence: 15

aaaacrccct aaaatccccg aaatcgccg 29

SEQ ID NO: 16

Length: 30

Type: DNA

Organism: Homo sapiens

Sequence: 16

aactcccrcc gaccccaacc ccgacgaccg 30

SEQ ID NO: 17

49

Length: 23
Type: DNA

Organism: Artificial Sequence

Feature:

Other Information: Description of Artificial

Sequence:Oligonucleotide

which binds bisulfite-converted human GST-Pi gene

Sequence: 17

aaacctaaaa aataaacaaa caa

23

SEQ ID NO: 18

Length: 23

Type: DNA

Organism: Artificial Sequence

Feature:

Other Information: Description of Artificial

Sequence:Oligonucleotide

which binds non-converted human GST-Pi gene

Sequence: 18

gggcctaggg agtaaacaga cag 23

SEQ ID NO: 19

Length: 25

Type: DNA

Organism: Artificial Sequence

Feature:

Other Information: Description of Artificial

Sequence:Oligonucleotide

which binds human GST-Pi gene

Sequence: 19

**5**0

cettteecte ttteccarrt cecca

25

SEQ ID NO: 20

Length: 25 Type: DNA

Organism: Artificial Sequence

Feature:

Description of Artificial Other Information:

Sequence:Oligonucleotide

which binds bisulfite-converted human GST-Pi gene

Sequence: 20

25 tttggtattt tttttcgggt tttag

SEQ ID NO: 21

Length: 25

Type: DNA

Organism: Artificial Sequence

Feature:

Other Information: Description of Artificial

Sequence:Oligonucleotide

which binds non-converted human GST-Pi gene

Sequence: 21

cttggcatcc tccccgggc tccag

25

SEQ ID NO: 22

Length: 26

Type: DNA

Organism: Artificial Sequence

Feature:

Other Information: Description of Artificial

Sequence:Oligonucleotide

which binds human GST-Pi gene

Sequence: 22

ggyagggaag ggaggyaggg gytggg

26

SEQ ID NO: 23

Length: 31

Type: DNA

Organism: Homo sapiens

Sequence: 23

ttatgtaata aatttgtata ttttgtatat g

31

SEQ ID NO: 24

Length: 25

Type: DNA

Organism: Homo sapiens

Sequence: 24

tgtagattat ttaaggttag gagtt

25

SEQ ID NO: 25

Length: 27

Type: DNA

Organism: Homo sapiens

Sequence: 25

aaacctaaaa aataaacaaa caacaaa

27

SEQ ID NO: 26

Length: 29

Type: DNA

Organism: Homo sapiens

52

Sequence: 26

aaaaaacctt tccctctttc ccaaatccc

29

SEQ ID NO: 27

Length: 27

Type: DNA

Organism: Homo sapiens

Sequence: 27

tttgttgttt gtttattttt taggttt

27

SEQ ID NO: 28

Length: 26

Type: DNA

Organism: Homo sapiens

Sequence: 28

gggatttggg aaagagggaa aggttt

26

SEQ ID NO: 29

Length: 24

Type: DNA

Organism: Homo sapiens

Sequence: 29

actaaaact ctaaacccca tccc

24

SEQ ID NO: 30

Length: 24

Type: DNA

Organism: Homo sapiens

Sequence: 30

aacctaatac taccttaacc ccat

PCT/AU99/00306 WO 99/55905

53

SEQ ID NO: 31

Length: 33 Type: DNA

Organism: Homo sapiens

Sequence: 31

aatcctcttc ctactatcta tttactccct aaa

33

SEQ ID NO: 32

Length: 29 Type: DNA

Organism: Homo sapiens

Sequence: 32

aaaacctaaa aaaaaaaaa aaacttccc

29

SEQ ID NO: 33 Length: 29

Type: DNA

Organism: Homo sapiens

Sequence: 33

ttggttttat gttgggagtt ttgagtttt 29

SEQ ID NO: 34

Length: 29

Type: DNA

Organism: Homo sapiens

Sequence: 34

29 ttttgtgggg agttggggtt tgatgttgt

SEQ ID NO: 35

Length: 29 Type: DNA

Organism: Homo sapiens

Sequence: 35

ggtttagagt ttttagtatg gggttaatt

29

SEQ ID NO: 36

Length: 20 Type: DNA

Organism: Homo sapiens

Sequence: 36

tagtattagg ttagggtttt

20

SEQ ID NO: 37

Length: 29
Type: DNA

Organism: Homo sapiens

Sequence: 37

aactctaacc ctaatctacc aacaacata

29

SEQ ID NO: 38

Length: 29
Type: DNA

Organism: Homo sapiens

Sequence: 38

caaaaaactt taaataaacc ctcctacca

29

SEQ ID NO: 39

Length: 32
Type: DNA

Organism: Homo sapiens

Sequence: 39

gttttgtggt taggttgttt tttaggtgtt ag

55

SEQ ID NO: 40

Length: 30

Type: DNA

Organism: Homo sapiens

Sequence: 40

gttttgagta tttgttgtgt ggtagttttt 30

SEQ ID NO: 41

Length: 30

Type: DNA

Organism: Homo sapiens

Sequence: 41

ttaatataaa taaaaaaat atatttacaa 30

SEQ ID NO: 42

Length: 34

Type: DNA

Organism: Homo sapiens

Sequence: 42

caacccccaa tacccaaccc taatacaaat actc 34

SEQ ID NO: 43

Length: 26

Type: DNA

Organism: Homo sapiens

Sequence: 43

ggttttagtt tttggttgtt tggatg 26

SEQ ID NO: 44

Length: 26

Type: DNA

WO 99/55905

PCT/AU99/00306

**5**6

Organism: Homo sapiens

Sequence: 44

tttttttgtt tttagtatat gtgggg

26

SEQ ID No: 45 Length: 30 Type: DNA

Organism: Homo sapiens

Sequence: 45

atactaaaaa aactattttc taatcctcta

30

SEQ ID NO: 46

Length: 29
Type: DNA

Organism: Homo sapiens

Sequence: 46

ccaaactaaa aactccaaaa aaccactaa

29

SEQ ID NO: 47 Length: 38

Type: DNA

Organism: Artificial Sequence

Feature:

Other Information: Description of Artificial Sequence: M13-human

GST-Pi oligonucleotide

Sequence: 47

tgtaaaacga cggccagtgg gatttgggaa agagggaa

38

SEQ ID NO: 48

Length: 38

Type: DNA

Organism: Artificial Sequence

Feature:

Other Information: Description of Artificial Sequence: M13-human

GST-Pi oligonucleotide

Sequence: 48

tgtaaaacga cggccagttg ttgggagttt tgagtttt

38

SEQ ID NO: 49

Length: 31

Type: DNA

Organism: Artificial Sequence

Feature:

Other Information: Description of Artificial Sequence: M13-human

GST-Pi oligonucleotide

Sequence: 49

tgtaaaacga cggccagtta gtattaggtt a

31

37

SEQ ID NO: 50

Length: 37

Type: DNA

Organism: Artificial Sequence

Feature:

Other Information: Description of Artificial Sequence: M13-human

GST-Pi oligonucleotide

Sequence: 50

tgtaaaacga cggccagtgt tttgagtatt tgttgtg

SEQ ID NO: 51

Length: 35

Type: DNA

58

Organism: Artificial Sequence

Feature:

Other Information: Description of Artificial Sequence: M13-human

GST-Pi oligonucleotide

Sequence: 51

tgtaaaacga cggccagtgt ttttagtata tgtgg 35

SEQ ID NO: 52 Length: 499 Type: DNA

Organism: Homo sapiens

Sequence: 52

SEQ ID NO: 53 Length: 316 Type: DNA

Organism: Homo sapiens

Sequence: 53

gggacctggg aaagaggaa aggetteece ggccagetge geggegaete eggggaetee 60 agggegeece tetgeggeeg aegeeegggg tgeageggee geeggggetg gggeeggeg 120 gagteegegg gacceteeag aagageggee ggegeegtga eteageaet gggeggageg 180 gggegggaee aeeettataa ggeteggagg eegegaggee ttegetggag tetegeegee 240 geagtetteg eeaceagtga gtaegeegg eeegegteee eggggatggg geteagaget 300

59

cccagcatgg ggccaa

316

SEQ ID NO: 54 Length: 603

Type: DNA

Organism: Homo sapiens

Sequence: 54

SEQ ID NO: 55 Length: 266 Type: DNA

Organism: Homo sapiens

Sequence: 55

getetgagea cetgetgtg ggeagtetet catcetteca egeacatect etteceetee 60 teccaggetg gggeteacag acageceet ggttggeeca teccagtga etgtggttg 120 ateaggegee cagteacgeg geetgeteee etecaceeaa eeceaggget etatgggaag 180 gaccageagg aggeageet ggtggacatg gtgaatgaeg gegtggagga eeteegetge 240 aaatacatet eecteateta eaceaa

SEQ ID NO: 56 Length: 287 Type: DNA

Organism: Homo sapiens

60

Sequence: 56

tcccctgct ctcagcatat gtggggegcc tcagtgcccg gcccaagctc aaggccttcc 60

tggcctcccc tgagtacgtg aacctcccca tcaatggcaa cgggaaacag tgagggttgg 120
ggggactctg agegggagc agagtttgcc ttcctttctc caggaccaat aaaatttcta 180
agagagctac tatgagcact gtgtttcctg ggacggggct taggggttct cagcctcgag 240
gtcggtggga gggcagagca gaggactaga aaacagctcc tccagca 287

SEQ ID NO: 57 Length: 524

Type: DNA

Organism: Homo sapiens

Sequence: 57

SEQ ID NO: 58 Length: 524 Type: DNA

Organism: Homo sapiens

Sequence: 58

tgttgtgatt tagtattggg gtggagtggg gtgggattat ttttataagg tttggaggtt 420 gtgaggtttt tgttggagtt ttgttgttgt agtttttgtt attagtgagt atgtgtggtt 480 tgtgtttttg gggatggggt ttagagtttt tagtatgggg ttaa 524

SEQ ID NO: 59 Length: 524 Type: DNA

Organism: Homo sapiens

Sequence: 59